Quercetin Ameliorates Zinc Oxide Nanoparticles-induced Impaired Testicular Functions by Regulating Inflammation, Oxidative, and Hormonal Status

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Abstract

Objective: Zinc oxide nanoparticles (ZnO NPs) have various applications, but concerns about their potential adverse effects on human health, particularly the male reproductive system, have led to toxicity. This study investigates the effects of quercetin, a natural compound with anti-oxidative and anti-inflammatory properties, on ZnO NP-induced testicular toxicity in male Swiss mice.

Method: In this study, 25 male mice were randomly allocated into 5 groups, each consisting of 5 mice (n=5). The groups were labelled as Control, Corn Oil, Quercetin, ZnO NPs, and ZnO NPs + Quercetin. The dosages administered were 100mg/kg for ZnO NPs and 20mg/kg for quercetin, relative to the weight of the animals. The treatments were conducted for 7 days following a two-week acclimatization period.

Results: Zinc oxide nanoparticles (ZnO NPs) were found to increase testicular TNF-α, indicating inflammation at the testes which may have resulted in a significant decrease in testosterone. Malondialdehyde (MDA), an oxidative stress marker was also found to be increased at the level of the testes, in the ZnO NPs group when compared to the control group. Treatment with quercetin, however, was able to reduce the levels of inflammatory and oxidative stress markers at the testes, and restore testosterone to control levels, demonstrating its potential ameliorative effects on testicular toxicity via anti-inflammatory and anti-oxidative properties.

Conclusion: The findings from this study indicated that Quercetin is a potential therapeutic agent against inflammation and oxidative stress in ZnO NPs-induced testicular toxicity.

Keywords: Zinc Oxide Nanoparticles, Quercetin, Inflammation, Oxidative stress, Testes

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Plain English Summary
This study looks at how quercetin, a natural substance that fights free radicals and inflammation, affects the testicular toxicity caused by Zinc Oxide nanoparticles (ZnO NPs) in male Swiss mice. Twenty-five male mice were randomly put into five groups of five mice each. Control, Corn Oil, Quercetin, ZnO NPs, and ZnO NPs + Quercetin were the names of the groups. The amounts given were 100 mg/kg for ZnO NPs and 20 mg/kg for quercetin, based on the weight of the animals for seven days. The results showed that zinc oxide nanoparticles raised testicular TNF-α levels. This means that the testes were inflamed, which may have caused the major drop in testosterone. Malondialdehyde (MDA), a sign of oxidative stress, was also found to be higher in the testes of the ZnO NPs group compared to the control group. However, treatment with quercetin was able to lower the levels of inflammatory and oxidative stress markers in the testes and bring testosterone levels back to normal. This study showed that quercetin may be able to help treat inflammation and oxidative stress in testicular damage caused by ZnO nanoparticles.

Introduction
Nanoparticles (NPs) are materials with dimensions between 1 to 100nm, offering unique properties like optical, reactivity, and toughness (1). They are suitable for various commercial, industrial, and domestic applications, including catalysis, imaging, medical, energy-based research, and environmental applications (1). Zinc oxide, a prominent metal nanoparticle, is increasingly common due to its unique properties in various industries (2). However, the increasing use of zinc oxide in consumer products increases the risk of exposure due to contamination from industrial processes and their easy entry into the systemic circulation through respiratory, gastrointestinal, and dermal pathways (3). These nanoparticles can cause toxicity in the reproductive system by disrupting sex hormone production, leading to oxidative stress, inflammation, DNA damage and cell death or apoptosis (4, 5).

Oxidative stress is a condition where the body's ability to detoxify or repair cellular damage is disrupted by the generation of reactive oxygen species (ROS), which are by-products of cellular metabolism (6). The testes, due to their high metabolic activity, lipid content, and reliance on unsaturated fatty acids, are particularly susceptible to oxidative stress (7). When ROS production exceeds antioxidant defences, oxidative stress leads to lipid peroxidation, protein damage, and inflammation (7). Inflammation, a complex immune response, can also cause testicular toxicity (8, 9). Inflammation involves the infiltration of immune cells into testicular tissue, releasing pro-inflammatory cytokines like TNF-α and IL-1β (10, 11). This can disrupt the normal testicular microenvironment, affecting spermatogenesis and causing oxidative stress. Chronic inflammation in the testes is associated with conditions like orchitis and epididymitis, which can result in testicular dysfunction and impaired sperm production (12). ZnO NP-induced increased testicular injury has been shown to induce oxidative stress and cytotoxicity in lab animals (5). In this scenario, antioxidant supplementation with quercetin becomes essential, and quercetin therapy is becoming more and more popular for treating repro-toxicant-related repro-pathologies (13, 14, 15).

Quercetin, a naturally occurring compound abundant in a wide range of fruits, vegetables, and herbs, has garnered considerable attention for its multifaceted health-promoting attributes (16). Notably, it exhibits remarkable anti-inflammatory properties and demonstrates the ability to shield the body from the harmful effects of diverse agents, such as heavy metals, pesticides, and nanoparticles, as demonstrated in the study conducted by Ay et al. in 2021 (17). The innate duality of zinc oxide nanoparticles (ZnO NPs) as promising technological innovations and potential health hazards underscores the urgency of investigating protective measures against their deleterious effects. This study sets out to examine the promising potential of quercetin as a mitigating agent in the context of testicular toxicity, specifically due to oxidative stress and inflammation induced by ZnO NPs.

Materials and Methods
Experimental model
Twenty-five (25) healthy male mice (20-25g) obtained from the animal house, Babcock University were kept at the animal house, Babcock University, Ilishan-Remo, Ogun State, Nigeria. The ethical approval for all handling of animals and protocols for the experiment used in the study was obtained from the Babcock University Health Research Ethics Committee, and this adheres to the international guidelines for using laboratory animals outlined in the Helsinki Declaration.

Animal care
The mice were housed at an average room temperature of 23 °C, with a relative humidity of 55% and a 12 h light / 12 h dark cycle and provided a standard diet and drinkable tap water ad libitum. The animals were acclimatized for two...
weeks at the animal house of the Department of Physiology, Babcock University.

**Experimental design**

Twenty-five (25) male Swiss mice were allocated into five groups (n = 5). The treatment group include the Control group (0.2mL of distilled water), the Corn Oil group (0.2mL of corn oil), Quercetin (20mg/kg), Zinc Oxide Nanoparticles (100mg/kg), Zinc Oxide Nanoparticles + Quercetin (100mg/kg+20mg/kg). The vehicles for the administration of Quercetin and Zinc Oxide nanoparticles were corn oil and distilled water respectively. The treatments were administered via oral cannula for seven days after 14 days of acclimatization.

**Drug administration**

Zinc oxide nanoparticles (ZnO NPs), quercetin, and corn oil were purchased from Sigma Aldrich, St. Louis, USA, Central Drug House (CDH) Limited, New Delhi, India, and Babcock Superstore, Ogun State, Nigeria respectively.

**Physicochemical characterisation of Zinc Oxide nanoparticle**

The dispersion protocol, morphology, and size distribution of Zinc Oxide Nanoparticles have been documented in our previous studies (18). The dispersion procedure and morphology assessment of Zinc nanoparticles were conducted in adherence to the methodology outlined by Georgantzopoulou et al. (2016) (19). Utilizing dynamic light scattering (DLS), the particle size distribution, zeta potential, and polydispersity indexes of ZnO NPs were determined, as detailed in the study by Cambier et al. (2018) (20).

**Sample collection**

After one week of treatment, blood was collected from each animal under ether anaesthesia through the retro-orbital sinus and centrifugated at 3000rpm for 15 minutes. The sera collected were used for hormonal analysis using the Enzyme-linked immunosorbent assay (ELISA) technique, and the animal was euthanized via cervical dislocation. The reproductive organs were extracted, separated from adhering tissues, and weighed using an electronic scale. The testes and epididymis were dissected for sperm analysis, biochemical assays, immunoassays, and histological examinations.

**Biochemical assays**

Malondialdehyde (MDA) assay was performed using the method described by Wallin et al. (21). The interaction of Malondialdehyde, a byproduct of lipid peroxidation, with thiobarbituric acid, produces a red species detectable at 535 nm. Superoxide Dismutase (SOD) was performed using Misra and Fridovich's (22) technique. The principle is based on the rapid auto-oxidation of the presence of superoxide anions, adrenaline in aqueous solution converts to adrenochrome. A spectrophotometer set to 420 nm was used to measure the concentration. Catalase (CAT) was determined using Aebi's (23) technique. Catalase (CAT) activity was measured as a reduction in H2O2 absorbance at 520 nm. Total antioxidant capacity (TAC) and reduced glutathione (GSH) were the methods by Ellman (24) and Degriff-Johnson (25) respectively. The Colorimetric method was used in the determination of testicular zinc levels using kits (Fortress Diagnostics Limited, United Kingdom).

**Hormonal and testicular TNF-α assay**

The Enzyme-linked immunosorbent assay (ELISA) technique was used to determine serum testosterone, luteinizing hormone, follicle-stimulating hormone and testicular tumour necrotic factor-alpha using kits (Calbiotech Inc. California). The procedures for the estimation of testosterone, luteinizing hormone and follicle-stimulating hormone were carried out according to the manufacturer instructions in the kits’ manual.

**Semen analysis**

Sperm motility

The sperm motility was performed immediately following the collection of epididymal fluid using Zemjanis’ (26) traditional technique. After dropping the epididymal fluid onto the slide, two drops of warm 2.9% sodium citrate were added to it. This was covered with a cover slip and examined under the microscope using an X40 objective in low light.

Sperm viability

The Eosin/Nigrosin stain was used for this examination. The epididymal sperm motility specimen was retrieved and the coverslip was rapidly removed. Two drops of Eosin/Nigrosin stain were applied to this, and a smear was produced and air-dried. The slides were observed under a microscope with an X40 objective. The live sperm cells were unstained; however, the dead sperm cells did. The % live/death ratio was calculated by counting 100 cells per slide (26).

Sperm count

The caudal epididymis was excised and homogenized in 5ml of normal saline, and the volume change was measured. Sperm count was determined using an improved Neubauer counting chamber. The counted sperm was expressed in million/ml suspension.
Histopathological examination of testes
The testis and hypothalamus were fixed in 10% buffered formalin, embedded in paraffin wax, and sectioned at 3μm. Sections were stained with Hematoxylin and Eosin (H&E) stain and mounted on slides for light microscopic inspection. To eliminate bias, the slides were reviewed by a histopathologist who was unaware of the treatment groups.

Statistical analysis
The data was expressed as Mean ± standard error of the mean (SEM). Analysis was carried out using one-way analysis of variance (one-way ANOVA) followed by the Bonferroni post hoc test for multiple comparisons in GraphPad Prism version 8 software. p < 0.05 was considered significant.

Results
The Effects of Zinc Oxide Nanoparticles and Quercetin on Testicular Zinc Levels
The results indicated a significant increase in zinc levels found in the testes of the nanoparticles group when compared to the negative control (p<0.001) as shown in Figure 1. Furthermore, Quercetin was also found to have mitigated the increased testicular zinc levels with a significant decrease in testicular zinc levels compared to the nanoparticles group (p<0.05).

Figure 1: The effects of nanoparticles and quercetin on testicular zinc levels
Data were expressed as Mean ± S.E.M (n = 5) and One-way ANOVA followed by the Bonferroni post hoc test was used for data analysis. ****p <0.0001 when compared with Control; #p < 0.05 when compared to Nanoparticles Group

The Effects of Zinc Oxide Nanoparticles and Quercetin on Testicular MDA Concentration
The result of this present study as shown in Figure 2 showed a significant increase in MDA concentration in the Zinc Oxide nanoparticles (ZnO NPs) group when compared to the control group. However, in the Nanoparticle+Quercetin group, there was a significant reduction in the mean testicular MDA concentration when compared to the Nanoparticle treatment group.

Figure 2: The effects of zinc oxide nanoparticles and quercetin on testicular malondialdehyde (MDA) concentration
Data were expressed as Mean ± S.E.M (n = 5) and One-way ANOVA followed by the Bonferroni post hoc test was used for data analysis. ****p <0.0001 when compared with Control; ####p < 0.0001 when compared to Nanoparticles Group

The Effects of Zinc Oxide Nanoparticles and Quercetin on Testicular Superoxide Dismutase (SOD) Concentration
The result as shown in Figure 3 showed that the nanoparticle treatment group showed a significant decrease in the mean testicular SOD concentration when compared to the control group. Furthermore, the Nanoparticle + Quercetin treatment group when compared to the Nanoparticle treatment group showed a significantly higher mean testicular SOD concentration.
The Effects of Zinc Oxide Nanoparticles and Quercetin on Testicular Superoxide Dismutase (SOD) Concentration

The result as shown in Figure 3 showed the mean testicular SOD concentration was significantly lower in the Nanoparticle treatment group when compared to the control group. Furthermore, there was no significant difference in the mean testicular SOD concentration between the Nanoparticle and the Nanoparticle + Quercetin treatment groups.

The Effects of Zinc Oxide Nanoparticles and Quercetin on Testicular Catalase (CAT) Concentration

The result as shown in Figure 4 showed the mean testicular CAT concentration was significantly lower in the Nanoparticle treatment group when compared to the control group. Furthermore, there was no significant difference in the mean testicular CAT concentration between the Nanoparticle and the Nanoparticle + Quercetin treatment groups.

The Effects of Zinc Oxide Nanoparticles and Quercetin on Testicular Glutathione (GSH) Concentration

The result as shown in Figure 5 showed a significant decrease in mean testicular GSH in the Nanoparticle treatment group when compared to the Control group. Furthermore, the Nanoparticle + Quercetin group did not show a significant difference when compared to the Nanoparticle only treated group.

The Effects of Zinc Oxide Nanoparticles and Quercetin on Testicular Total Antioxidant Capacity (TAC)

The result as shown in Figure 6, showed that the mean testicular Total Antioxidant Capacity (TAC) of the nanoparticle-treated group showed a significant decrease when compared to the control group.
Control group. However, the Nanoparticle+Quercetin showed increased levels of the mean testicular Total Antioxidant capacity (TAC) when compared to the Nanoparticle treatment group.

![Figure 6. The Effects of Zinc Oxide Nanoparticles and Quercetin on Testicular Total Antioxidant Capacity (TAC)](image)

Data were expressed as Mean ± S.E.M (n = 5) and One-way ANOVA followed by the Bonferroni post hoc test was used for data analysis. ****p <0.0001 when compared with Control; ##p < 0.01 when compared to Nanoparticles Group

**The Effects of Zinc Oxide Nanoparticles and Quercetin on TNF-α**

A significant increase in TNF-α was indicated in the nanoparticles group when compared to the control (p<0.0001) as illustrated in Figure 7, however, quercetin was shown to ameliorate the effects of the nanoparticles as the co-administration group showed no significant difference in TNF-α when compared to the control group.

![Figure 7: The effects of nanoparticles and quercetin on TNF-α](image)

Data were expressed as Mean ± S.E.M (n = 5) and One-way ANOVA followed by Bonferroni post hoc test was used for data analysis ***p <0.001 when compared with Control; ****p <0.0001 when compared with Control; ####p < 0.0001 when compared to Nanoparticles Group

**The Effects of Zinc Oxide Nanoparticles and Quercetin on Follicle-Stimulating Hormone**

The results as illustrated in Figure 8 show no significant changes in serum FSH levels in both the nanoparticles group and the co-administration group to the control group, as well as no significant alteration in serum FSH levels of the co-administration group when compared to the nanoparticles group.

![Figure 8: The effects of nanoparticles and quercetin on follicle-stimulating hormone](image)

Data were expressed as Mean ± S.E.M (n = 5) and One-way ANOVA followed by the Bonferroni post hoc test was used for data analysis

**The Effects of Zinc Oxide Nanoparticles and Quercetin on Luteinizing Hormone**

The results as illustrated in figure 9 showed no significant difference among the groups when compared to the control or the nanoparticle group.
The Effects of Zinc Oxide Nanoparticles and Quercetin on Serum Testosterone Levels

The results as presented in Figure 10 indicate a significant decrease in testosterone levels in the Nanoparticle-treated animals (p <0.0001). Moreover, quercetin was found to ameliorate the deleterious effects of Nanoparticles, with its testosterone levels showing no significant difference compared to the control. As shown in the graph below, there was a significant difference in serum testosterone levels in the co-administration group compared to the Nanoparticle-treated group (p<0.0001).

![Figure 10: The effects of nanoparticles and quercetin on serum testosterone levels](image)

Data were expressed as Mean ± S.E.M (n = 5) and One-way ANOVA followed by the Bonferroni post hoc test was used for data analysis. ****p <0.0001 when compared with Control; ####p < 0.0001 when compared to Nanoparticles Group

The Effects of Zinc Oxide Nanoparticles and Quercetin on Testosterone-LH Ratio

The results as shown in Figure 11 indicate a significant decrease in the testosterone-LH ratio when compared to the Control group (p<0.0001). Furthermore, quercetin was able to ameliorate the deleterious effects of the nanoparticles in the nanoparticles and quercetin group co-administration group (p<0.0001) when compared to the control.

![Figure 11: The effects of nanoparticles and quercetin on testosterone-LH ratio](image)

Data were expressed as Mean ± S.E.M (n = 5) and One-way ANOVA followed by the Bonferroni post hoc test was used for data analysis. ****p <0.0001 when compared with Control; ####p < 0.0001 when compared to Nanoparticles Group

The Effects of Zinc Oxide Nanoparticles and Quercetin on Sperm Motility

The result as shown in Table 1 reveals that the treatment groups showed no significant difference in sperm motility, viability and count when compared to the control group. Moreover, the result also showed no significant difference in sperm motility, viability and count between the Nanoparticle treatment group and those of the Nanoparticle + Quercetin treatment group.
Table 1: The effects of Zinc Oxide Nanoparticles and Quercetin on Sperm motility, viability and count

<table>
<thead>
<tr>
<th>Treatment Group</th>
<th>Sperm Motility (%)</th>
<th>Sperm Viability (%)</th>
<th>Sperm Count (million/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>86.00 ± 2.45</td>
<td>90.40 ± 1.91</td>
<td>104.20 ± 6.14</td>
</tr>
<tr>
<td>Corn Oil</td>
<td>81.00 ± 2.45</td>
<td>86.00 ± 1.87</td>
<td>116.60 ± 4.84</td>
</tr>
<tr>
<td>Quercetin</td>
<td>85.00 ± 1.58</td>
<td>91.00 ± 1.87</td>
<td>103.20 ± 7.43</td>
</tr>
<tr>
<td>ZnO NPs</td>
<td>81.00 ± 3.32</td>
<td>90.00 ± 1.58</td>
<td>104.40 ± 5.40</td>
</tr>
<tr>
<td>ZnO NPs + Quercetin</td>
<td>86.00 ± 1.87</td>
<td>91.00 ± 1.87</td>
<td>113.40 ± 5.87</td>
</tr>
</tbody>
</table>

Data were expressed as Mean ± S.E.M (n = 5) and One-way ANOVA followed by the Bonferroni post hoc test was used for data analysis.

The Effects of Zinc Oxide of Nanoparticles and Quercetin on Histology of the Testes

Results show that intoxication with ZnO resulted in degeneration of testicular morphology characterized by decentralised lumen, with necrotic Leydig cells and pyknotic Sertoli cells. Furthermore, the testes showed hypertrophy with germinal epithelium that is disarrayed and fragmented. However, treatment of Quercetin alongside ZnO significantly normalized the testicular cytoarchitecture as the cellular assortments in this group show a well-centralized lumen. Moreover, Leydig cells are properly arranged and surrounded by connective tissues in the interstitial space and the germinal epithelium was properly arranged.

Figure 12: The effects of zinc oxide of nanoparticles and quercetin on the histology of the testes

C - Control, CO - Corn Oil, QUE - Quercetin, and ZnO NPs – Zic Oxide nanoparticles

Discussion

This study aimed to investigate the anti-oxidative and anti-inflammatory effects of quercetin administration against testicular toxicity induced by zinc oxide nanoparticles (ZnO NPs). We observed a significant increase in testicular zinc concentration in the group treated with ZnO NPs when compared to the control group. This elevation in zinc levels can be attributed to the dissolution of ZnO NPs, leading to the release of zinc ions, a phenomenon documented in previous research (27). Interestingly, in the ZnO NPs+Quercetin group, we noted a significant difference in testicular zinc concentration when compared to the group treated with zinc oxide nanoparticles alone. This observation aligns with the known metal-chelating properties of quercetin, as reported in prior studies (28). These findings suggest that quercetin may exert its influence by interacting with zinc ions, potentially chelating or sequestering them to some extent.

Oxidative stress arises from an imbalance between pro-oxidant and antioxidant substances within a biological system, as highlighted in research by Pizzino et al. (29). This imbalance triggers lipid peroxidation, leading to the formation of aldehydes and related compounds such as Malondialdehyde (MDA), as described in the study by Moldogazieva et al. (30). MDA levels have been widely utilized as a reliable indicator for assessing oxidative stress, as demonstrated in studies by Altun et al. (31) and Ghonimi et al. (32). In our study, we observed significantly elevated testicular MDA levels in the group exposed to zinc oxide nanoparticles (ZnO NPs) when compared to the control group. However, in the ZnO NPs+Quercetin group, a notable reduction in testicular MDA concentration was evident in comparison to the ZnO NPs group.
This finding suggests that quercetin effectively mitigated the oxidative stress induced by ZnO NPs within the testes. This result aligns with the findings reported by Li et al. (33), who demonstrated a decrease in MDA concentrations in a rat model of Acute Myocardial Infarction following quercetin administration. In a study by Xu et al. (34) on silk worms exposed to ZnO NPs, SOD and CAT levels initially increased, but a decline was observed after 36 hours. The study hypothesized that excess superoxide ions might inhibit SOD activity, indirectly affecting CAT. Our current study aligns with this hypothesis, as it revealed a significant decrease in mean testicular SOD and CAT concentrations in the Nanoparticle treatment group. Interestingly, the Nanoparticle+Quercetin group displayed a notable increase in mean testicular SOD levels, indicating that quercetin had an ameliorative effect. However, quercetin was unable to restore CAT concentrations, which remained significantly reduced.

Testicular Total Antioxidant Capacity (TAC), an indicator of testicular antioxidant activity, reveals a significant decrease in the Zinc Oxide Nanoparticles treatment group. Conversely, quercetin ameliorated this effect in the Zinc Oxide Nanoparticle + Quercetin treatment group. This suggests that co-administration of quercetin may have restored the total antioxidant capacity to levels akin to the normal physiological state. These findings can be corroborated by the research conducted by Ziamajidi et al. in 2023 (35), which demonstrated the successful mitigation of ZnO NP-induced oxidative stress by well-known antioxidants such as Vitamins A, C, and E. This further underscores the remarkable antioxidant potential of Quercetin and its ability to counteract the oxidative effects triggered by ZnO NPs.

Multiple studies (36, 37, 38) have examined the impact of zinc oxide nanoparticles (ZnO NPs) on hormone levels. These studies collectively show that ZnO NPs tend to raise follicle-stimulating hormone (FSH) and testosterone levels while leaving luteinizing hormone (LH) levels relatively unchanged. For instance, a 2020 study on male NMRI rats revealed increased serum testosterone and FSH levels with intra-peritoneal ZnO NP administration (37). Conversely, another study on mice reported a dose-dependent decrease in testosterone levels following oral ZnO NP administration (36). Our study, however, aligns with this later finding, indicating a significant reduction in testosterone levels without significant changes in FSH and LH levels. These discrepancies may be due to variations in exposure duration, nanoparticle dosage, animal models, and administration routes. The decrease in testosterone indicated in this study could be a result of inflammation at the Leydig cells of the testes. This is corroborated by the results of this study which shows a significant increase in the concentration of testicular TNF-α, a pro-inflammatory cytokine in the nanoparticle-treated group when compared to the control. Quercetin however was shown to ameliorate the effect of zinc oxide nanoparticles on testosterone levels. Several plausible mechanisms may underlie these observed effects. First, quercetin has demonstrated metal-chelating properties in multiple studies, enabling it to bind with zinc within cells, and aiding in the elimination of zinc from the body (39). This aligns with our findings of a significant reduction in testicular zinc oxide levels in the groups treated with quercetin. Additionally, quercetin's ameliorative effects might be attributed to its anti-inflammatory mechanisms. Our analysis of tumour necrotic factor (TNF-α) levels revealed a significant increase in the nanoparticle-treated groups. TNF-α, an inflammatory cytokine produced during acute inflammation by macrophage (40), has been linked to increased expression due to zinc oxide nanoparticles, signifying the presence of cellular inflammation (41). Our study supports this by showing significantly elevated TNF-α levels in the group treated solely with zinc oxide nanoparticles compared to the control, indicating testicular inflammation. Quercetin demonstrated anti-inflammatory properties in our study, as evidenced by its ability to mitigate the potentially pro-inflammatory effects induced by zinc oxide nanoparticles in the testes. This was highlighted by a significant reduction in TNF-α levels in the co-administration group compared to the nanoparticle group.

Also, in this study, the observed decrease in serum testosterone levels relative to serum luteinizing hormone (Testosterone-LH ratio) in the nanoparticle-treated group compared to the control holds important implications. It signifies a reduction in testosterone production from the Leydig cells, while LH secretion remained unaffected. This imbalance, characterized by low testosterone levels concerning LH, is associated with primary hypogonadism or primary testicular failure (42). Such an outcome may be attributed to potential damage to the Leydig cells, impairing their ability to produce testosterone despite the continued secretion of LH by the anterior pituitary. This finding is consistent with the increased levels of the inflammatory marker TNF-α in the nanoparticle-treated groups compared to the control, indicating the presence of inflammation in the testes, as demonstrated in our study. Moreover, other research studies have also reported heightened inflammatory responses at various sites, including the testes, following nanoparticle exposure (43, 44).
The spermatogenesis cycle in mice typically spans around 35 days, as indicated by Perrard et al. (45). During this process, germ cells transform spermatozoa. In our study, the treatment period was limited to 7 days, which falls short of completing one full cycle of spermatogenesis. This shortened exposure duration may explain the absence of significant alterations in sperm parameters, including motility, viability, and count, as revealed in our semen analysis. This underscores the importance of considering the timing and duration of treatments when assessing their impact on sperm parameters and reproductive outcomes in animal studies.

Lastly, histological examination of the testes revealed degeneration of testicular morphology due to ZnO NP administration. This finding can be corroborated by the results of Mozaffari et al. (2015) (44). However, in the group receiving a combination of ZnO NPs and Quercetin, the histological analysis revealed a normal cytoarchitecture, indicating the ameliorative effects of Quercetin on the alterations induced by ZnO NPs.

**Conclusion**

Quercetin in this study was shown to demonstrate its anti-inflammatory and anti-oxidative properties against zinc oxide nanoparticles inducing testicular toxicity by restoring serum testosterone levels.

**List of Abbreviations**

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
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<tbody>
<tr>
<td>MDA</td>
<td>Malondialdehyde</td>
</tr>
<tr>
<td>SOD</td>
<td>Superoxide dismutase</td>
</tr>
<tr>
<td>CAT</td>
<td>Catalase</td>
</tr>
<tr>
<td>GSH</td>
<td>Glutathione peroxidase</td>
</tr>
<tr>
<td>TAC</td>
<td>Total antioxidant capacity</td>
</tr>
<tr>
<td>ZnO NPs</td>
<td>Zinc oxide nanoparticles</td>
</tr>
<tr>
<td>TNF-α</td>
<td>Tumour necrotic factor – alpha</td>
</tr>
<tr>
<td>FSH</td>
<td>Follicle-stimulating hormone</td>
</tr>
<tr>
<td>LH</td>
<td>Luteinizing hormone</td>
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</tbody>
</table>

**Declarations**

*Ethics approval and consent to participate*

Ethical approval was obtained from Babcock University Health Research Ethics Committee (BUHREC), with registration number 320/24.

*Consent for Publication:*

All the authors gave their consent for the publication of the work under the Creative Commons Attribution Non-Commercial 4.0 license. Otherwise, all copyright ownership including all rights incidental thereto is conveyed to the journal when published.

*Availability of data and materials*

The study data is available upon request to the corresponding author.

**Competing interests**

The authors declare that there are no conflicts of interest.

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The authors did not receive any funding from funding agencies.

**Authors’ contributions**

Conceptualization, OBO, GOO, EWU and EGO.; methodology, OBO, OMO, CEO and TVO.; validation, OBO, PGO and EGO; formal analysis, OBO, CEO, EGO and TK; investigation, CEO. TVO; resources, OBO, CEO. TVO and EWU; data curation, OBO and CEO; writing—original draft preparation, OBO, ADA, TK and TFA; writing—review and editing, OMO, ADA, PGO, GOO; supervision, OBO; project administration, ADA, CEO and TVO; funding acquisition, OBO, CEO and TVO. All authors have read and agreed to the published version of the manuscript.

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